Chronic Hypertension Leads to Neurodegeneration in the TgSwDI Mouse Model of Alzheimer’s Disease

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Abstract—Numerous epidemiological studies link vascular disorders, such as hypertension, diabetes mellitus, and stroke, with Alzheimer’s disease (AD). Hypertension, specifically, is an important modifiable risk factor for late-onset AD. To examine the link between midlife hypertension and the onset of AD later in life, we chemically induced chronic hypertension in the TgSwDI mouse model of AD in early adulthood. Hypertension accelerated cognitive deficits in the Barnes maze test ($P<0.05$ after 3 months of treatment; $P<0.001$ after 6 months), microvascular deposition of β-amyloid ($P<0.001$ after 3 months of treatment; $P<0.05$ after 6 months), vascular inflammation ($P<0.05$ in the dentate gyrus and $P<0.001$ in the dorsal subiculum after 6 months of treatment), blood–brain barrier leakage ($P<0.05$ after 3 and 6 months of treatment), and pericyte loss ($P<0.05$ in the dentate gyrus and $P<0.01$ in the dorsal subiculum after 6 months of treatment) in these mice. In addition, hypertension induced hippocampal neurodegeneration at an early age in this mouse line (43% reduction in the dorsal subiculum; $P<0.05$), establishing this as a useful research model of AD with mixed vascular and amyloid pathologies. (Hypertension. 2015;66:00-00. DOI: 10.1161/HYPERTENSIONAHA.115.05524.)

Key Words: blood–brain barrier ■ cerebral amyloid angiopathy ■ hypertension

Alzheimer’s disease (AD) is the most common form of dementia, and there is no universal cause or treatment to delay or stop its progression. Late-onset AD often occurs without the contribution of known genetic risk factors and results in memory loss, irritability, and eventually death.1–2 Cerebral amyloid angiopathy (CAA), the deposition of β-amyloid (Aβ) along vessel walls in the central nervous system, is observed in >94% of patients with AD.3–5

In addition to CAA, researchers have observed numerous ultrastructural and functional changes within the AD microvasculature. Alterations in every cellular component of the neurovascular unit (NVU), the tightly regulated network of cells that couples neuronal energy demands to modulation of blood flow, have been observed in patients with AD. Furthermore, both endothelial cells and pericytes degenerate in brain capillaries in patients with AD.4,5 Loss of these cell types has detrimental effects on blood–brain barrier (BBB) integrity, as well as neuronal perfusion and function.6–8 Astrocyte endfeet, which ensheath brain capillaries, help regulate capillary blood flow, and maintain the extracellular milieu, are swollen in the presence of CAA.6,7 The direct cause of these cellular abnormalities has not been determined although Aβ is toxic to neurons and other cell types in vitro.9 In addition, the presence of Aβ deposits among the cells of the NVU could interfere with signaling between cell types, alter cellular health and function, and result in reduced flow-mediated dilation, an indicator of vessel reactivity, also observed in AD brains.10

Thus, CAA may contribute to the reduced neurovascular coupling reported in multiple AD mouse lines.10–13

More than 30% of AD cases exhibit cerebrovascular pathology in addition to CAA,14 and the frequent coincidence of stroke and AD suggests that the cerebrovascular changes that occur during AD progression compromise vascular integrity and function.15 Clinical evidence suggests that cardiovascular risk factors, such as hypertension, are linked to AD onset.14

In non-AD individuals, hypertension induces pathological changes in the brain, including impaired cerebral autoregulation, vascular remodeling, cerebral microbleeds, and cerebral atrophy.16–18 Given the vasoactive properties of Aβ,19 it is unclear whether midlife hypertension is an early symptom of the vascular pathology present in AD or whether it contributes to the onset of the disease. It is possible that elevated blood pressure (BP) during midlife compromises vascular integrity and leads to cellular, basement membrane, and BBB damage. Given the prevalence of cardiovascular risk factors in middle-aged individuals and the relevance of these risk factors to AD susceptibility, we induced chronic hypertension in an AD mouse model. We examined behavioral, cellular, and ultrastructural changes to determine the effect of chronic hypertension before AD onset on disease pathogenesis.
shown). The Barnes maze24 was used to compare the cognitive levels of CD31 or treated animals (Figure 1A). There were no differences in exhibited a significant increase in BP compared with water-
during testing/analysis, respectively. L-NAME–treated mice for 3 or 6 months and were 6 to 7 or 9 to 10 months of age
changes in this mouse line (not shown). Mice were treated before we observed any dramatic AD-related pathological
posed to AD, we induced hypertension at 3 to 4 months of
18,22,23 Because TgSwDI mice are predis-
individuals with midlife hypertension before indications of
was designed to mimic the pattern of hypertension
and increase survivorship during long-term treatment.20,21 Our
TgSwDI and WT mice. L-NAME treatment alone was used to
reduce the likelihood of hemorrhage and increase survivorship during long-term treatment.20,22 Our protocol was designed to mimic the pattern of hypertension that predisposes humans to late-onset AD, which occurs in individuals with midlife hypertension before indications of cognitive decline.20,22,23 Because TgSwDI mice are predisposed to AD, we induced hypertension at 3 to 4 months of age, which corresponds to early adulthood in humans and before we observed any dramatic AD-related pathological changes in this mouse line (not shown). Mice were treated for 3 or 6 months and were 6 to 7 or 9 to 10 months of age during testing/analysis, respectively. L-NAME–treated mice exhibited a significant increase in BP compared with water-treated animals (Figure 1A). There were no differences in levels of CD31 or α-smooth muscle actin between normoten-
sive and hypertensive brains, suggesting that hypertension did not affect vessel number in the brains of these animals (not shown). The Barnes maze24 was used to compare the cognitive abilities of normotensive and hypertensive WT and TgSwDI mice. After 3 months of L-NAME treatment, TgSwDI mice performed significantly worse during the 5-day probe trial compared with water-treated TgSwDI mice and all WT mice (Figure 1B), indicating that chronic hypertension affects cogni-
tion after only 3 months, well before significant cognitive impairment is evident from the transgene alone. TgSwDI mice treated with L-NAME for 6 months exhibited a similar trend (not shown). After 6 months of treatment, hypertensive TgSwDI mice performed worse during Barnes maze training, taking significantly longer to find the escape hole compared with all other groups (Figure 1C), an effect that was not observed in TgSwDI mice after 3 months of treat-
ment. There were no differences in baseline locomotor activity between any of the groups (not shown). Interestingly, this cohort of water-treated TgSwDI mice (9–10 months of age) did not have cognitive deficits relative to WT mice in this test, although chronic hypertension was able to induce significant cognitive dysfunction.

### Chronic Hypertension Induces Vascular Amyloid Deposition in TgSwDI Mice

To determine whether increased levels of Aβ in the hippocampus were responsible for the cognitive deficits observed in our hypertensive TgSwDI mice, we used thioflavin-S to detect fibrillar Aβ. We costained sections with an anti-collagen IV antibody, which recognizes the basement membrane of capillaries. After quantifying the total length of collagen IV-positive capillaries in the dorsal subiculum (DS), we quantified the length of vessels costained by thioflavin-S to calculate the percent of capillary CAA. We found that Aβ in the DS of normotensive TgSwDI mice (Figure 2A and 2C) was both microvascular and parenchymal. Hypertensive TgSwDI mice (Figure 2B and 2E), however, had fewer parenchymal deposits and exhibited significantly more microvascular CAA (Figure 2C and 2F). Although localization of Aβ deposition in the brains of normotensive and hypertensive TgSwDI mice was different, thioflavin-S intensity overall was not (not shown), suggesting that the amount of Aβ in these groups was similar. Immunostaining
with anti-Aβ40- and 42-specific antibodies revealed no significant difference in levels of either peptide between normotensive and hypertensive TgSwDI mice, though both were slightly elevated in the hypertensive group (not shown), suggesting that both peptides contribute to CAA load in these mice. Immunoelectron microscopy confirmed that Aβ deposition appeared as finger-like clusters around microvessels (Figure 2G).

Abundant Microglia Surround Microvessels of Hypertensive TgSwDI Mice

Because activated astrocytes and microglia are often abundant in Aβ-laden tissue, we examined glial markers in normotensive and hypertensive TgSwDI and WT brains. Although levels of astrocyte markers were similar in both TgSwDI groups (not shown), the microglial marker ionized calcium–binding adapter molecule 1 was upregulated in hippocampal subregions of hypertensive TgSwDI mice compared with those of normotensive mice (Figure 3A and 3D versus Figure 3B and 3E). Ionized calcium–binding adapter molecule 1 expression was significantly elevated in water-treated TgSwDI mice compared with water-treated WT mice and was even higher in L-NAME–treated TgSwDI mice (Figure 3C and 3F). The pattern of ionized calcium–binding adapter molecule 1 staining in the dentate gyrus (DG) appears patchy (Figure 3B), likely due to the presence of larger parenchymal plaques in this region, compared with the DS, where the staining appears vascular (Figure 3E). We examined Aβ-laden microvessels by electron microscopy and confirmed that CAA is often surrounded by infiltrating microglia (Figure 3G, M).

Hypertension Disrupts Tight Junctions and Decreases BBB Integrity in TgSwDI Mice

Because the BBB is disrupted in patients with AD and mouse models, we examined the BBB integrity to determine whether hypertension affects this feature of AD pathology. Under normal conditions, tight junctions, which form the basis of the BBB, appear continuous and lay flat, preventing diffusion of blood components into the brain (Figure 4A). However, the tight junctions in samples from hypertensive TgSwDI mice appeared to be breaking off into the capillary lumen and lifting slightly from the endothelial cell layer (arrowheads in Figure 4B–4D), providing an opportunity for BBB leakage. Given these structural alterations, we examined tissue for the presence of blood components that could enter the brain if BBB integrity were compromised. Albumin was elevated in the DS and DG of hypertensive TgSwDI brains when compared with normotensive TgSwDI brains after only 3 months of treatment, indicating that BBB integrity was compromised because of hypertension (Figure 4E–4H).

Chronic Hypertension Results in the Loss of Neurons and Pericytes in TgSwDI Brains

Under normal conditions, many AD mouse models, including TgSwDI mice, do not exhibit neurodegeneration, a

![Figure 2](image-url)
hallmark of AD pathology. Because of the accelerated time course of other pathologies in L-NAME–treated TgSwDI mice, we examined hippocampal subregions for neuronal death. Neurodegeneration was not evident in normotensive TgSwDI mice (Figure 5A and 5C), but significant neuronal loss was observed after short-term L-NAME treatment (3 months) and at only 6 to 7 months of age (Figure 5B and 5C). Normotensive TgSwDI mice at 9 to 10 months of age began to exhibit subtle signs of neuronal loss, making hypertension-induced cell death at this age less dramatic (Figure 5D–5F).

Pericyte loss occurs in severely affected patients with AD33 and in older TgSwDI mice12; so, we examined the levels of platelet-derived growth factor receptor β to determine whether the expression of this pericyte marker is altered in our hypertensive groups. After 6 months of L-NAME treatment, hypertensive TgSwDI mice exhibited a significant decrease in platelet-derived growth factor receptor β staining in both the DG and DS (Figure 5G and 5J versus Figure 5H and 5K; Figure 5I and 5L). Taken together, these results suggest that neuronal and pericytic loss can occur at a much earlier age when there is concomitant hypertension.

**Discussion**

Under normal conditions, hypertension induces pathological changes within the cerebral vasculature, resulting in impaired autoregulation, microbleeds, and lacunar infarcts, as well global downstream changes, such as white matter lesions and atrophy.16,34–39 In fact, these changes within the cerebral vasculature of hypertensive individuals also occur in patients with AD in the absence of hypertension.40–42 In addition, midlife hypertension is a significant risk factor for the development of AD later in life.44 Hypertension may initiate vascular damage before the onset of AD, allowing symptoms to be more pronounced and progress more quickly. In addition, compromised vessels may be more vulnerable to the deleterious effects of Aβ. For example, because hypertension results in reduced BBB integrity (Figure 4), blood components may enter the brain before large-scale accumulation of Aβ occurs. These blood components may serve as a seed for Aβ deposition and increase vascular inflammation (Figure 3), resulting in cellular damage and the release of toxic molecules.

The hippocampus coordinates memory consolidation and spatial navigation and is one of the first regions in the brain to suffer damage in AD.45 The DG receives all hippocampal inputs and passes them through the hippocampus proper (CA1-4) to the subiculum, which projects to the entorhinal cortex.44 Thus, proper functioning of the DG and subiculum is essential to memory consolidation and spatial navigation. The subiculum may also be involved in the spread of Aβ, as lesions in this region inhibit plaque formation in other brain areas.45

We found that the location, but not abundance, of Aβ deposits is altered in hypertensive TgSwDI mice, which also exhibit early and dramatic cognitive decline. Our findings suggest that CAA contributes more to cognitive decline than parenchymal plaques. We also observed that microvascular Aβ adheres to capillary basement membranes that normally bind cells of the

![Figure 3](image-url)

**Figure 3.** Hypertensive TgSwDI mice display increased vascular microgliosis. Microgliosis was examined in tissues using an anti–ionized calcium–binding adapter molecule 1 (iba-1) antibody (green) and vessels were stained with an endothelial cell–specific anti-CD31 antibody (red). Compared with water-treated TgSwDI (A and D) and wild-type (WT; not shown) groups, the dentate gyrus (DG; B) and dorsal subiculum (DS; E) of hypertensive TgSwDI brains exhibited significantly more iba-1 staining after 6 months of treatment (C and F; **P<0.01 normotensive TgSwDI vs normotensive WT; ***P<0.001 normotensive vs hypertensive TgSwDI; scale bar, 100 μm; n=6–9 per group). G. The infiltration of microglia (M) was observed by electron microscopy of brain capillaries in Hypertensive TgSwDI samples. Capillaries laden with β-amyloid (*) were surrounded by infiltrating microglial nuclei (scale bar, 2 μm). L indicates lumen.
levels, hypertensive TgSwDI mice without an overall increase in Aβ deposition were often lifted or fragmented (arrowheads in B–D). The plasma protein albumin, which was present at low levels in the dorsal subiculum (DS) of normotensive TgSwDI brains (E), was significantly enriched in the DS (F–H; *P<0.05) and dentate gyrus (DG; not shown) of hypertensive TgSwDI mice after 3 months of L-NAME treatment. Compared with normotensive wild-type (WT) mice, hypertensive WT mice also exhibited significant leakage of albumin from the vasculature into the DS after 3 months of L-NAME treatment (H; *P<0.05; scale bar, 1 μm (A) and 20 μm (E); n=4–6 per group).

NVU together. Therefore, CAA may act as a physical barrier to cell signaling at the NVU. Moreover, CAA seems to obstruct binding of astrocytic endfeet and contribute to pericyte loss (Figure 5). Both astrocytes and pericytes are involved in the recruitment of blood flow during neuronal activity, and damage to these cell types in hypertensive TgSwDI mice may impair neurovascular coupling.

Aβ is thought to be deposited as CAA because of failed clearance across the BBB and along perivascular spaces. Because L-NAME treatment results in increased CAA in TgSwDI mice without an overall increase in Aβ levels, hypertension may have a detrimental effect on either or both of these clearance pathways. L-NAME inhibits endothelial nitric oxide synthase and thereby prevents the production of the vasodilator nitric oxide. Alternatively, it could be that nitric oxide–mediated vasodilation in normotensive mice is required for effective clearance of Aβ across the BBB or along perivascular drainage pathways.

The neuronal loss we observed in hypertensive TgSwDI mice could result from reductions in blood flow and impaired function of NVU support cells that either degenerate (pericytes; Figure 5) or are chronically activated (microglia; Figure 3). Because brain atrophy is associated with hypertension in humans and hypertensive WT mice exhibit a trend toward reduced neuron number, it is possible that vascular changes that occur during hypertension in WT mice, such as BBB leakage (Figure 4), affect neuronal health. However, the changes specific to hypertensive TgSwDI mice, such as increased microgliosis (Figure 3) and severe CAA (Figure 2), are likely to contribute in some way. In addition to the cytotoxic effects of Aβ itself, activated microglia are known to produce cytotoxic molecules, which may damage neurons.

We used L-NAME to induce hypertension because of its long-term tolerability and ease of administration to large cohorts of mice. L-NAME treatment resulted in elevated BP similar to other treatment types but circumvented drawbacks of other surgical and chemical techniques. Surgical wounds are susceptible to infection or injury, often requiring animals to be singly housed and excluded from behavioral experiments. Techniques requiring infusion pumps are shorter in duration, hindering the implementation of chronic hypertension, and more severe treatment regimes, such as the pairing of L-NAME with other molecules, resulted in seizures and intracerebral hemorrhage in our mice (not shown). Therefore, long-term L-NAME treatment was the best suited to mimic chronic hypertension in humans, which can last decades.

Although models of comorbid hypertension and AD are limited, our results are consistent with what has been observed in other models. Similar to our findings, surgically induced hypertension in WT mice results in cognitive deficits and subtle amyloid pathology, whereas angiotensin-II infusion in APPPS1 mice results in a more rapid onset of AD pathology. However, it is possible that longer-term L-NAME–induced hypertension exacerbates previously observed pathologies, such as cognitive decline (Figure 1) and microvascular Aβ deposition (Figure 2), and reveals new neuropathologies not previously observed, such as microglia activation (Figure 3), BBB leakage (Figure 4), and pericytic and neuronal loss (Figure 5).

Studies linking hypertension with AD define midlife as the time between the ages of 40 and 64 years. Midlife hypertension is associated with an increased risk of developing AD, but late-life hypertension (typically defined as age ≥65) does not have this association. Importantly, studies linking hypertension with AD examine BP before onset of AD symptoms. Given that untreated TgSwDI mice develop some AD pathology 6 months after onset of AD symptoms (Figures 2 and 3) and cognitive decline 10 to 12 months (not shown), we initiated hypertension at an age corresponding to early adulthood developmentally, to maximize the duration of hypertension before the onset of AD symptoms. Although midlife hypertension is a significant risk factor for AD and treatment alleviates this risk, dramatic reduction in BP often occurs at later stages of AD, after which point antihypertensives are deleterious to cognitive function. Hypertension may compromise vascular integrity during midlife and lead to cellular, basement membrane, and BBB damage. However, after the onset of AD symptoms, low BP may aggravate the brain hypoperfusion already present in AD because of other types of vascular damage.

We found that hypertension dramatically accelerated various features of AD pathology in our model. Microvascular Aβ deposition was increased in hypertensive TgSwDI mice without an increase in overall Aβ deposition (Figure 2), suggesting that Aβ is recruited to capillaries in hypertensive individuals. Because we showed that L-NAME–treated...
TgSwDI mice have deficits in cognitive function (Figure 1B and 1C), our results suggest that capillary CAA is detrimental to cognitive function relative to other forms of deposited Ab, an idea that has been proposed by others in the field.\textsuperscript{54,55} It is possible that the deposition of microvascular Ab compromises the survival of NVU cells, such as pericytes and endothelial cells. Although others have shown pericyte loss in TgSwDI mice at 18 months of age,\textsuperscript{12} we found that chronic hypertension induced pericyte loss significantly earlier (9–10 months of age; Figure 5G–5L). Furthermore, neuronal loss has never been reported in this transgenic model, yet hypertensive TgSwDI mice demonstrated significant neurodegeneration in a subregion of the hippocampus (Figure 5A–5F). Moreover, not only did hypertension result in quantifiable neuronal loss, but it did so much earlier (6–7 months of age) than the few other mouse lines that exhibit this feature of AD.\textsuperscript{56} At 6 to 7 months of age, the first indications of AD (Ab deposition, neuroinflammation, and cognitive decline) are often just becoming apparent in other AD mouse models.\textsuperscript{56–58} Our finding is notable because neuronal loss is widely accepted as a hallmark of AD yet does not occur in many commonly used AD models.\textsuperscript{39} If hypertension indeed accelerates AD pathogenesis, then a reasonable hypothesis is that neuronal loss is a downstream effect of the abundant microvascular amyloid pathology observed in TgSwDI mice. Pairing hypertension with genetic predisposition to AD may represent a more complete model of AD than the traditional AD mouse model.

Figure 5. Hypertensive TgSwDI mice exhibit early neuron and pericyte loss. Anti-NeuN antibody was used to identify neurons in the dorsal subiculum (DS) of normotensive (A and D) and hypertensive (B and E) TgSwDI mice. C. Reduced NeuN intensity was observed in hypertensive TgSwDI mice after 3 months of L-NAME treatment (\(P<0.05\); n=3–9 per group). F. Because NeuN intensity was slightly decreased in normotensive TgSwDI animals after 6 months of treatment because of advanced age (9–10 months of age), there was no longer a significant difference between groups. Although NeuN intensity appeared reduced in the DS of hypertensive wild-type (WT) mice compared with normotensive WT mice after 3 and 6 months of treatment, neither change was significant (C and F). Similarly, anti–platelet-derived growth factor receptor β (PDGFRβ) antibody was used to examine pericyte coverage of vessels in normotensive (G and J) and hypertensive (H and K) TgSwDI mice. I and L, PDGFRβ levels were significantly reduced in hypertensive TgSwDI mice in the dentate gyrus (DG) and DS compared with control groups after 6 months of treatment (\(P<0.05\), \(\ast P<0.01\); scale bar, 50 μm; n=5–9 per group).

Our results suggest that hypertension has a significant effect on the onset and progression of AD pathology. Treating hypertension in midlife may be an effective strategy for reducing the likelihood of AD onset later in life, as suggested by numerous epidemiological studies.\textsuperscript{17,53} Vascular damage because of untreated hypertension results in loss of vascular tone and hypotension in older individuals. Treatment with antihypertensives after the onset of cognitive symptoms in patients with AD exacerbates cognitive decline,\textsuperscript{60} possibly because of further reduction in neurovascular coupling. If blood flow were reduced to areas of the brain requiring energy substrates, then low BP would only exacerbate this effect and damage already-compromised neurons and NVU components.\textsuperscript{61}

**Perspectives**

We show that hypertension accelerates cognitive decline and other AD pathologies—CAA, neuroinflammation, BBB leakage, pericyte loss, and neurodegeneration—in transgenic mice predisposed to AD. Because many individuals with late-onset AD experience some form of midlife cardiovascular-related disease, such as hypertension,\textsuperscript{32} our mouse model of mixed vascular and amyloid pathologies may be more relevant for studying human AD pathophysiology than a mouse model of AD alone.

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Disclosures

None.

References


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Hypertension induces brain β-amyloid accumulation, cognitive impairment, and memory deterioration through activation of receptor for advanced glycation end products in brain vasculature. 

Summary

Because hypertension accelerates AD pathogenesis, our results suggest that neuronal loss could be part of the disease trajectory in this and possibly other mouse lines if mouse models were longer lived. Given that (1) hypertensive TgSwDI mice exhibited neuronal loss and many other AD hallmarks and (2) many individuals with late-onset AD experience some form of midlife cardiovascular–related disease, our mouse model of mixed vascular and amyloid pathologies may be more relevant for studying human AD pathophysiology than a mouse model of AD alone.